

ge-related macular degeneration (AMD) is the most common cause of visual impairment among the elderly in developed countries, and its prevalence is thus increasing as the population ages; however, treatment options remain limited because the etiology and pathogenesis of AMD are incompletely defined. Recently, much progress has been made in gene discovery and mechanistic studies, which clearly indicate that AMD involves the interaction of multiple genetic and environmental factors. The identification of genes that have a substantial impact on the risk for AMD is not only facilitating the diagnosis and screening of populations at risk but is also elucidating key molecular pathways of pathogenesis. Pharmacogenetic studies of treatment responsiveness among patients with the "wet" form of AMD are increasingly proving to be clinically relevant; pharmacogenetic approaches hold great promise for both identifying patients with the best chance for vision recovery as well as tailoring individualized therapies.



■ Yuhong Chen^{1,2}, Matthew Bedell², and Kang Zhang^{2,3}

¹Department of Ophthalmology and Vision Science, Eye and ENT Hospital, Fudan University, Shanghai, 200031, China

²Institute for Genomic Medicine and Shiley Eye Center, University of California at San Diego, La Jolla, CA, 92093-0838

³Department of Ophthalmology, West China Hospital, Sichuan University, 610041, China

Introduction

Age-related macular degeneration (AMD) is the most common cause of visual impairment among elderly individuals in the developed world, occurring primarily in persons over the age of 50 (1–7). It is estimated that 1.75 million people in the United States suffer from advanced AMD, and 7.3 million people are affected with intermediate AMD, which represents increased risk for development of advanced disease (8). AMD thus poses a major public health problem with significant economic and social impact.

AMD is an abnormality of the retinal pigment epithelium (RPE) that leads to photoreceptor degeneration of the overlying central retina, or macula, and loss of central vision (9–10) (Figure 1). The macula (Figure 2) is 5–6 mm in diameter, and at its center is the fovea, responsible for greatest visual acuity. Early AMD is characterized by subretinal deposits, known as *drusen*, that mea-

sure greater than 60 µm and hyper- or hypo-pigmentation of the RPE. Intermediate AMD is characterized by the accumulation of focal or diffuse drusen measuring greater than 125 µm and hyper- or hypo-pigmentation of the RPE (Figure 2). Advanced AMD can be classified into either of two categories. The first of these comprises geographic atrophy (GA; i.e., dry, or non-exudative, AMD), which is characterized by a sharply delineated area of REP atrophy measuring at least 175 um along one dimension and including visible choroidal vessels (Figure 2C). The alternative form of the advanced disease is choroidal neovascularization (CNV; i.e., wet, or exudative, AMD), which may involve some or all of the following: subretinal neovascular membranes; subretinal fluid, exudates, and hemorrhages (Figure 2D); pigment epithelial detachment (PED; see Figure 1B); and subretinal/intraretinal scarring. Advanced AMD can result in loss of central visual acuity and lead to severe and permanent visual impairment and blindness. Whereas dry

AMD accounts for 80–90% of all cases of advanced disease, more than 90% of AMD patients with severe loss of central vision manifest CNV (11).

The etiology and pathophysiology of AMD are poorly understood. Drusen, which are the histological markers of AMD, are yellowish extracellular deposits of lipid, protein, lipoprotein, and cellular debris that accumulate between the RPE and Bruch's membrane or within Bruch's membrane (12–15). Drusen can include complement components and modulators (4, 16–21) and the serine protease HTRA1 (22). The cause of drusen may be linked to one or more of the following key processes that are implicated in AMD: 1) increased outer segment turnover; 2) impaired activity/function of the RPE; 3) free radical/oxidative damage; 4) aging and degeneration of elements of Bruch's membrane (e.g., collagen and elastin); 5) reduced clearance of material from Bruch's membrane into choriocapillaries; and 6) deleterious immune system activa-

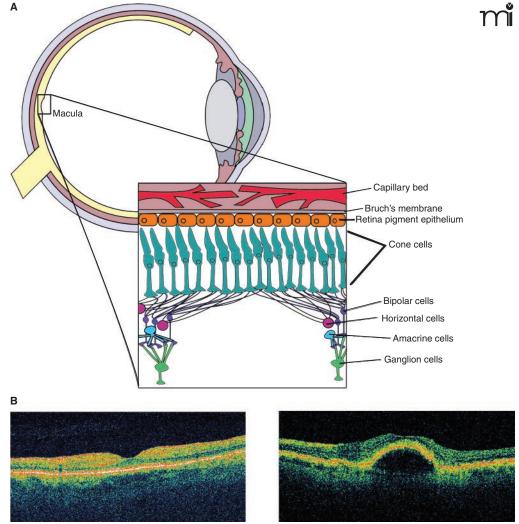


Figure 1. The retina revealed. A. Diagrammatic representation of the retina. Inset shows the several types of neuronal cells within the macula, the retinal pigment epithelium (RPE), and choroid. B. Two optical coherence tomography images of a normal (left) and AMD retina with pigmented epithelium detachment (PED) (right).

tion (23). Specific alterations in Bruch's membrane composition that have been suggested to cause drusen include excessive lipid deposition and protein cross linking as well as impaired permeability to nutrients.

GENETIC AND ENVIRONMENTAL RISK FACTORS

Although advancing age is the greatest risk factor associated with the development of AMD, environmental and lifestyle factors may significantly affect individual risk. Smoking is an important, modifiable factor that has been consistently associated with a twofold increased risk for developing AMD (odds ratios ranging from 1.8 to 3) (24). Oxidative stress and antioxidant depletion have been implicated in retinal damage from smoking, although the precise mechanism in AMD remains unclear (25–27). Other factors that have been reported to influence risk for AMD include sunlight exposure, alcohol consumption, increased plasma fibrinogen levels, diet, hypertension, body mass index (BMI), and iris color (24, 27–31). Regarding diet, antioxidants such as carotenoids, zinc, and vitamins A and E may provide a protective benefit against AMD.

Epidemiological studies have demonstrated differences in the prevalence of AMD based on ethnicity, with prevalence among Caucasians being greater than that among non-white groups (32–33). Such ethnic differences may reflect genetic as well as environmental risk factors. AMD has a significant genetic component (34–37), as several twin studies have shown significantly higher concordance rates between monozygotic twins as compared to concordance between dizygotic twins (38–42). Familial aggregation studies from general populations (43–44) and tertiary eye care centers (45–46) reveal that family members of individuals with AMD are at increased risk (2.4- to 19.8-fold) for developing the disease relative to individuals with no family history.

GENETIC STUDIES OF AMD

Gene association studies reveal that multiple genes may be associated with AMD (Table 1). An association between AMD and the gene that encodes apolipoprotein E (APOE) has been reported, through multiple studies (47–51), such that the ε 2 allele increases risk, whereas the $\varepsilon 4$ allele has a protective effect. In a screen for sequence variation among the gene family that encodes the fibulin (FBLN) glycoproteins, Stone and colleagues detected missense mutations in 1.7% of 402 AMD patients (52). Seven different missense mutations of the FBLN5 gene have been discovered in individuals examined from eight families affected by AMD with a distinctive mutation corresponding to each of the affected individuals (53). Evidence from genetic association studies in conjunction with the demonstration of complement deposition in the retina and choroid has implicated toll-like receptor-(TLR)-3 and -4 in the development of certain cases of AMD (53, 54). Most recently, two large genome-wide association studies have established another two susceptibility loci. A functional promoter variant of the hepat-

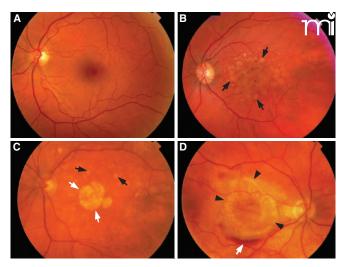


Figure 2. Fundus images from patients representing several AMD subtypes. A. Normal macula. B. Macula with confluent soft drusen (black arrows), a hallmark of early AMD. C. Macula of dry AMD with soft drusen (black arrows) and geographic atrophy GA (white arrows). D. Macula of choroidal neovascularization (CNV) or wet AMD with subretinal hemorrhage (black arrows).

ic lipase–encoding gene (*LIPC*) was found to be strongly associated with advanced AMD, corroborating the involvement of lipid pathways in AMD development (55). Additionally, a locus near the gene for metallopeptidase inhibitor 3 (*TIMP3*), which is involved in degradation of the extracellular matrix, was found to be associated with AMD (56). Other candidate genes include *C3*, *C2/CFB*, *VEGFA*, *ABCA4*, *ERCC6*, and *CX3CR1* (Table 1).

The most convincing evidence for the genetic contribution to AMD is the identification of major disease susceptibility alleles on chromosomes 1q32 (which includes the gene that encodes complement factor H [CFH]) and chromosome 10q26 (which includes PLEKHA1, hypothetical gene LOC387715, and HTRA1). Specifically, risk of developing AMD is associated with an allele of CFH in which a histidine residue is encoded in place of a tyrosine residue at amino acid position 402 (16, 57, 58). The increased risk ranges between 2- to 4-fold for heterozygote carriers and 3- to 7-fold for homozygotes. In addition, multiple other polymorphisms, many of which are in non-coding regions of CFH or in nearby genes encoding other complement factors, demonstrate equal or stronger association with disease susceptibility than does the CFH Y402H variant (16, 59). Importantly, no single polymorphism could account for the entire contribution of CFH to disease susceptibility. Rather, multiple polymorphisms defined a set of four common haplotypes (two of which promoted disease susceptibility and two of which were seemingly protective) and several comparatively rare haplotypes (all of which were associated with increased disease susceptibility) (60, 61). At 10q26, containing genes PLEKHA1, LOC387715, and HTRA1 (62-65), a single nucleotide polymorphism (SNP) in the promoter of the HTRA1 gene was associated with a population attributable risk of 49.3% and a 10-times greater risk of developing CNV (22, 66).

Table 1. Genetic Loci Associated with AMD

Gene symbol (name)	Function	Position ^b	Variant	Odds Ratios (OR) ^c	References
ABCA4 (ATP-binding cassette, sub-family A, member 4)	Photoreceptor-specific expression; transport of N-retinylidene-PE across the outer segment disc membrane	1p22	rs 1800553, rs 1800555	Conflicting results: OR _{hetero} =5.0 OR _{hetero} =2.8 no association	(102, 103)
APOE (Apolipoprotein E)	Lipid and cholesterol transport	19q13	rs429358, rs7412	Conflicting results: ε2 OR _{homo} =1.046 ^d ; ε4 OR _{homo} =0.35-0.53, 0.847 ^d ; no association	(103–105)
ARMS2/LOC387715° (Age-related maculopathy susceptibility 2)	Unknown; gene prod- uct is localized to mitochondrial outer membrane	10q26	rs10490924, in/	OR _{homo} =8.59	(27, 103, 106)
HTRA 1 ^a (High temperature requirement A1)	Trypsin-like serine protease	10q26	rs11200638	OR _{homo} =6.92 ^d , 7.46 ^d	(107, 108)
C2/CFB (Complement 2/ Complement factor B)	Regulation of comple- ment activation	6p21	rs9332739 (c2), rs4151667(CFB), rs641153 (CFB)	OR _{hetero} =0.32-0.40	(59, 105)
C3 (Complement 3)	Innate immunity (alternative complement pathway activator, classical pathway component)	19p13	rs2230199, rs1047286	OR _{homo} =1.93-3.91	(74, 75, 105)
CFH ^a (Complement factor H)	Inhibitor of alternative complement pathway	1q32	rs1061170	OR _{homo} =6.32 ^d	(27, 109)
CFHR1/CFHR3 (Complement factor H-related 1, 3)	Unknown, possible overlapping function with <i>CFH</i>	1q31-q32	84K bp deletion	OR _{homo} =0.29	(110)
CX3CR1 (Chemokine [C-X3-C motif] receptor 1)	Inflammation (chemokine receptor)	3p21	rs3732378	OR _{homo} =1.98-2.70	(111, 112)
ERCC6 (Excision-repair cross-complementing, group 6)	DNA repair	10q11	rs3793784	OR _{homo} =1.6	(113)
TLR3 (Toll-like receptor 3)	Innate immunity (tar- gets+ viral dsRNA)	4q35	rs3775291	OR _{homo} = 0.44-0.61	(54, 114, 115)
TLR4 (Toll-like receptor 4)	Innate immunity (bacterial endotoxin receptor)	9q32-q33	rs4986790	Conflicting results: OR _{hetero} =2.65; no association	(53, 54, 114, 115)
VEGFA (Vascular endothelial growth factor A)	Angiogenesis	6р12	rs833069, rs1413711	Conflicting results: OR _{homo} =5.29 OR _{homo} =2.40 no association	(116–119)

molecular interventions

 ^a Strong association.
 ^b Chromosomal position.
 ^c OR_{homo'} homozygous odds ratio for risk allele; OR_{hetero'} heterozygous odds ratio for risk allele.
 ^d Metaanalysis.

The preeminence of the *HTRA1* SNP in accounting for the association of 10q26 to disease was subsequently established also in a Utah population; however, *LOC387715* manifested very high association as well, suggesting that the AMD risk with regard to 10q26 may, similar to the scenario in the *CFH* region, reflect linkage as a function of alternative haplotypes. When disease-associated alleles exist within both the *HTRA1* and *CFH* loci, the population attributable risk for AMD is estimated to be 71.4% (22).

PATHOGENIC MECHANISMS IN AMD

The complement activating genes *CFH*, *CFB*, and *C2*, as well as the serine protease family gene *HTRA1*, have the strongest associations with AMD and offer a glimpse into possible pathogenic mechanisms. *LOC387715* is also highly associated. But because it has no known protein function, its role in pathogenesis cannot be adequately hypothesized. *CFH* binds and inactivates *C3b*, an important complement protein, and this process contributes to the selectivity of innate immune responses (*67*). Domain 7 in *CFH* has been shown to contain the site of binding specificity that allows *CFH* to recognize heparin or sialic acid on host cells (*68*).

The AMD-associated Y402H polymorphism discussed earlier happens to occur within domain 7 of CFH (16), possibly reflecting that the binding specificity of CFH is altered in such a way as to preclude recognition of host cell surface molecules and the corresponding inactivation of bound C3b. Accordingly, the Y402H substitution would leave host cells, particularly RPE and choroid-associated cells, vulnerable to complement-mediated degradation, which is consistent with the observation that CFH is expressed in drusen (16). Whether or not CFH variants result in drusen formation or merely accumulate within drusen is not known; however, with CFH expressed in drusen, its accumulation results in increased risk of RPE and choroidal cell degradation, threatening the overlaying retina. The end result would be photoreceptor

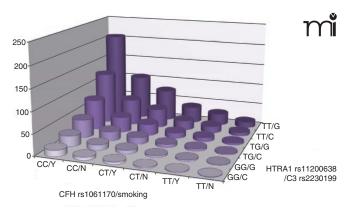


Figure 3. AMD risk profile as a function of disease-associated genes and smoking status. Joint effect of specific alleles of *CFH* (rs1061170, Y402H), *HTRA1* (rs11200638), *C3* (rs2230199) and smoking status were shown by odds ratios calculated by a logistic regression model, adjusted for age, gender, and body mass index (BMI).

degeneration and, ultimately, AMD. Other CFH, CFB, and C2 variants could act through similar pathways.

The AMD-associated promoter polymorphism observed in HTRA1 presumably alters a binding element (recognized by transcription factors AP2 and SRF) and thereby potentiates HTRA1 expression (22, 62). HTRA1 is a serine protease and a key modulator of proteoglycans degradation in the extracellular matrix (69). HTRA1 proteolytic activity permits other degradative enzymes, such as collagenases and matrix metalloproteinases, to access their respective substrates (70). Similar to complement factors, HTRA1 is expressed in drusen of AMD patients (22). Excessive accumulation of HTRA1 in drusen could compromise Bruch's membrane integrity, thereby allowing expansion of choroidal capillaries and resulting in neovascular AMD. HTRA1 is also a potent inhibitor of transforming growth factor- β , which is involved in extracellular matrix formation and angiogenesis (71) and thus represents another potential pathway to AMD. Alternatively, HTRA1-mediated destabilization of Bruch's membrane could contribute to RPE atrophy and GA.

RISK PREDICTION MODELS

The key to establishing the mechanisms of pathogenesis in AMD will depend on an understanding of the interplay between the genetic and environmental factors that have been implicated in the disease. The development of risk models that consider this interplay facilitates the identification of patients at greatest risk for developing AMD or progressing through the various stages of the disease. Risk models have been developed to predict the prevalence and incidence of AMD by duly considering multiple gene variations [i.e., in *CFH* (57, 72, 73), *HTRA1/LOC387715* (22, 66), *C2* (59), *CFB* (59), and *C3* (74, 75)] in concert with environmental, ocular, demographic, behavioral, and treatment factors (76).

Multiplicative models demonstrate that each genetic variant noted above is independently associated with AMD, and when patients carry high-risk variants on both of the major susceptibility genes, the risk of developing AMD increases tremendously (Figure 3). The discrimination accuracy of predictive models is limited to known factors, accounting for only about 40-60% (27, 65, 77-79) of the genetic risk for developing AMD in Caucasians (after adjustment for other genetic and environmental factors). Recently, sensitivity has been improved to 88% in an AMD predictive model through the inclusion of biomarkers of complement components and activation fragments (i.e., Ba, C3a, C3d, C5a, and factor D) (80, 81); however, specificity still remains around 73%. (Here, "sensitivity" is the probability that the model correctly identifies AMD-affected individuals, whereas "specificity" is the probability that the model correctly identifies healthy individuals.) In order to predict the occurrence and progression of AMD precisely enough to be used clinically for diagnosis and prognosis, it is crucial that we identify additional genetic or environmental contributors for incorporation into predictive models.

Early identification of high-risk individuals could allow

clinicians to better preserve patient vision through targeted surveillance and therapeutic intervention. High-risk individuals would also benefit from targeted education regarding healthy lifestyles. Smoking is the most important and obvious environmental factor that patients could eliminate to decrease their risk for AMD. Although the combined risk for AMD increases with age, regardless of genetic background, smoking increases the probability of developing AMD by 10–15%. Furthermore, carrying even one risk allele of *CFH* advances the potential age of disease onset by 10 years, regardless of smoking status (Figure 4).

Implications of Genotype in the Treatment of AMD

The etiology and pathophysiology of AMD are becoming increasingly better characterized as the number of genetic factors implicated in the pathogenesis of AMD increases. Understanding the role of genetics in AMD allows both direct and indirect tailoring of therapies to target specific molecules and pathways. However, studies must first prove that therapeutic differences correlate with genotypes before clinical practice recommendations can be changed. Currently, there is a small but growing body of evidence showing that an individual's genetic profile may influence existing therapies for AMD.

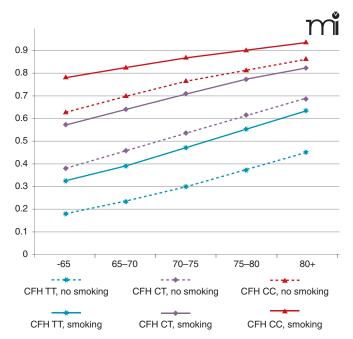


Figure 4. Lifelong risk for developing AMD. Probabilities of developing advanced AMD were estimated by evaluating genotypes of *CFH* (rs2274170) and smoking status. Low risk, median risk, and high risk associated with *CFH* genotypes were coded as blue, purple, and red, respectively. Solid lines represent smoking group and dashed lines represent nonsmoking group.

GENOTYPE AND RESPONSE TO PHOTODYNAMIC THERAPY

Photodynamic therapy (PDT) is a treatment for AMD-related CNV. Intravenous administration of the photosensitizer verteporfin, followed by activation with a non-thermal laser, initiates photochemical reactions, involving singlet oxygen and reactive oxygen intermediates, that damage endothelial cells lining the CNV; the therapeutic rationale is thus to prevent thrombosis and selective occlusion within CNV vessels. However, the efficacy of PDT treatment is limited. Although several ocular predictive factors, such as baseline visual acuity and CNV size at presentation, have been considered to explain the individually variable efficacy of PDT, they have only weakly contributed to effective optimization of PDT. Evaluations of the potential role of common AMD-associated gene variants in determining CNV responsiveness to PDT have been inconsistent. Brantley and colleagues found that visual acuity in PDT-treated individuals with the CFH TT genotype was significantly worse than that of PDT-treated TC and CC genotypes (82). Goverdhan and colleagues found the degree of visual loss following PDT to be significantly higher in the CFH CC genotype (83). Two other studies (84-85) reported no significant association between CFH genotype and responsiveness to PDT. No effect upon PDT responsiveness has been associated with particular LOC387715 or HTRA1 variants (86). Instead, two SNPs affecting the CRP gene and two SNPs in the VEGF gene (see below) demonstrated significant association with response to PDT (85, 87). Moreover, peculiar polymorphisms in genes that encode coagulation balance factors (e.g., factor V, prothrombin, and factor XIII-A) as well as in the MTHFR gene have been correlated to the degree of post-PDT benefit in Caucasian patients with neovascular AMD (88-90). Although patient cohort size was limited in these retrospective studies, the correlations identify an opportunity to optimize patient eligibility criteria for PDT (Table 2).

GENOTYPE AND RESPONSIVENESS TO ANTI-VEGF Treatment

Vascular endothelial growth factor (VEGF) is a major molecular mediator of neovascularization and is present in CNV membranes in wet AMD. VEGF inhibitors have been used to successfully treat exudative AMD and have become clinical standards. Ranibizumab is a humanized antibody fragment that targets VEGF-A and all of its biologically active degradation products. Phase 3 clinical trials showed that ranibizumab can stabilize vision in greater than 90% of patients and significantly improve vision over a two-year period in one-quarter to one-third of patients with neovascular AMD; the drug was approved by the FDA for treatment of neovascular AMD in 2006 (91–92). Bevacizumab is an FDA-approved anti–VEGF-A antibody for treatment of metastatic colorectal and breast cancer. Retrospective case series have demonstrated efficacy and safety of off-label bevacizumab for treating neovascular AMD (93). The ongoing National Eye Institute-(NEI)-sponsored comparison of

AMD treatment trials will compare ranibizumab with bevacizumab, administered monthly, for the treatment of neovascular AMD in 1200 patients over a two-year period.

An investigation of the effects of CFH and HTRA1/LOC387715 genotypes on the responsiveness of neovascular AMD to intravitreal bevacizumab revealed that visual acuity improved in only 10.5% of treated patients who were homozygous for the CFH Y402H (CC) genotype, whereas visual acuity improved in 53.7% of treated patients who possessed CFH TC and TT genotypes (P = 0.004). In fact, mean visual acuity worsened, from 20/206 to 20/341, among the patient population who possessed the CC genotype (P = 0.016). On the other hand, the LOC387715 genotype did not appear, after adjusting for age, pretreatment visual acuity, and lesion size, to affect patient responsiveness to bevacizumab (P=0.18) (94). Because both bevacizumab and ranibizumab target the same VEGF-A molecules, it is expected that the same pharmacogenetic relationship should exist for ranibizumab. Indeed, a retrospective analysis of 156 patients with exudative AMD who had been treated with intravitreal ranibizumab monotherapy and followed for nine months (95), indicates that patients with the CFH (CC) genotype tended to require more injections of drug than did other patients, which is indicative of a potential pharmacogenetic relationship between CFH genotype and ranibizumab treatment outcome (Table 2).

Table 2. Pharmacogenetic Studies of Treatment Outcome in AMD

Intervention	Gene/locus	Variants	Results	
Photodynamic therapy	CFH	rs1061170 (Y402H)	Controversial: Outcome for CC genotype lagged CT and TT (83); outcome for TT was poorer (82); no genotype association (84, 85).	
	LOC387715	rs10490924 (A69S)	No significant genotype association (86).	
	VEGF	rs699947, rs2146323	Anatomic outcome was strongly linked to SNPs (87).	
	CRP	rs2808635, rs876538	Positive response was significantly associated with both variants (85).	
	HTRA 1	rs11200638	No significant association (86).	
	FV FII	G1691A G20210A	Better outcome associated in patients carrying both genetic variants (88, 89).	
	MTHFR	C677T	Better outcome associated with variant (88–90).	
	FXIIIA	G185T	Poorer outcome associated with variant (88–89).	
Intravitreal bevacizumab	CFH	rs1061170 (Y402H)	CC genotype responded significantly worse than TC and TT (94); CC genotype more likely to require re-injection (95).	
	LOC387715	rs10490924 (A69S)	No significant association (94).	
Antioxidants and zinc	CFH	rs1061170 (Y402H)	TT genotype responds better than CC (98).	
	LOC387715	rs10490924 (A69S)	No significant association (98).	

GENOTYPE AND NUTRITIONAL SUPPLEMENTATION

In 2001, the Age-Related Eye Disease Study (AREDS), a large, multicenter, double-masked, placebo-controlled clinical trial, established that a combination of zinc and antioxidants (β-carotene, vitamin C, and vitamin E) produced a 25% reduction in development of advanced AMD (over a five-year period) and a 19% reduction in severe vision loss in individuals determined to be at high risk of developing the advanced forms of the disease. Use of these oral supplements has become standard practice in the US and remains the only therapy for intermediate and advanced dry AMD (96-97). The Study also revealed that CFH Y402H genotype could influence the effect of zinc in the supplementation regime. Specifically, the greatest benefit was seen in those individuals with the low-risk CFH genotype, among whom the rate of AMD progression was lowered by approximately two-thirds. In contrast, AREDS-type supplements seem to have less impact on those with the high-risk CFH genotype (98). Moreover, an interaction (P= 0.004) was observed in the AREDS treatment groups taking zinc when compared with the groups taking no zinc, but not in groups taking antioxidants compared with those taking no antioxidants (P = 0.59). These findings suggest that the genotype-treatment interaction in participants with CFH TT genotype may be related primarily to the zinc component of the supplements. Genetic screening could

thus help to identify those individuals at high risk for developing advanced AMD and could benefit patients who are the most likely to respond to supplements (Table 2).

Conclusions

Great strides have been made in the past five years in identifying genes associated with AMD, although much work is yet needed to understand the complex molecular genetics of this disease. Future endeavors, building on current scientific successes. should include: 1) detailed phenotyping to more accurately describe populations; 2) large-scale population studies allowing for better interpretation of mixed findings; and 3) functional studies of AMD-associated genes in both in vitro and in vivo systems.

Future studies need to put more emphasis on the harmonization of descriptions (e.g., primary features, age of onset, environmental factors) of AMD cases. Large datasets can then be generated to allow stratification of general phenotypes for genome-wide linkage analysis and association studies into specific phenotypes, such as GA and wet AMD, or primary features, such as soft drusen. There may, for example, be genes that confer risk for GA, or for exudative AMD, or for both together. Certain genes may help determine the subset of early AMD that progresses to advanced AMD. Analysis of quantitative trait loci, which assumes that the genes that confer risk for the late form are the same genes as for the early form, may lose power if such genes are distinct. Further defining subphenotypes with respect to environmental factors could reveal gene-environment interactions that would otherwise go undetected.

Accurate phenotype descriptions, standardized across multiple populations, may lead to high-density marker sets and supply result consistency and statistical power in analyzing further association and linkage studies. Many of the genes in Table 1 have been strongly associated in one or more population studies but not in other populations. High-density genotyping arrays will enhance consistency by providing richer genetic descriptions of populations studied.

Finally, functional studies are necessary to demonstrate that AMD-associated genetic variants can indeed lead to biological changes. Techniques from cell transfection to genetically engineered animals will allow researchers to examine the functional roles of multiple alleles in AMD onset and progression. Genetically engineered animals are ideal for not only understanding the pathophysiology of AMD, but also for discovering possible therapeutic interventions. The ability to reproduce genetic findings in cell and animal studies is a necessary step in attributing true disease association to specific genetic variants.

In this way, the burgeoning field of pharmacogenetics will enable clinicians to tailor pharmacotherapy to a patient's specific genetic variations. Identification of genetic risk variants will be useful in identifying high-risk populations for early treatment. Examples of diseases for which a genetic profile has already assisted in treatment include statin treatment in cardiovascular disease (99), imatinib treatment for chronic myeloid leukemia (100), and warfarin dosing with respect to cytochrome P450 CYP2C9 genotypes (101). As one of the most well-characterized, late-onset, complex diseases in terms of clinical features and underlying genetic and environmental influences, AMD is an excellent disease to which the principles of personalized medicine can be applied. Early detection of patients with high-risk genotypes and implementation of smoking cessation should significantly reduce the age of onset and severity of AMD. Furthermore, genotype-based customizing of treatment of neovascular AMD by anti-VEGF therapy will allow clinicians to identify optimal responders to maximize treatment benefit and reduce costs and side effects. doi:10.1124/mi.10.5.4

Acknowledgments

This work was supported by the National Institutes of Health [Grants R01EY14428, R01EY18660]. We further wish to acknowledge support from Research to Prevent Blindness, the Ruth and Milton Steinbach Fund, Ronald McDonald House Charities, the Macular Vision Research Foundation, Burroughs Wellcome Fund Clinical Scientist Award in Translational Research, and West China Hospital of Sichuan University.

References

- Penfold PL, Killingsworth MC, and Sarks SH (1985) Senile macular degeneration: the involvement of immunocompetent cells. *Graefes Arch Clin Exp Ophthalmol* 223:69–76.
- Klein R, Klein BE, and Linton KL (1992) Prevalence of age-related maculopathy. The Beaver Dam eye study. Ophthalmology 99:933–943.
- Vingerling JR, Dielemans I, Hofman A, Grobbee DE, Hijmering M, Kramer CF, and de Jong PT (1995) The prevalence of age-related maculopathy in the Rotterdam Study. Ophthalmology 102:205–210.
- Hageman GS, Luthert PJ, Victor Chong NH, Johnson LV, Anderson DH, and Mullins RF (2001) An integrated hypothesis that considers drusen as biomarkers of immune-mediated processes at the RPE-Bruch's membrane interface in aging and age-related macular degeneration. *Prog Retin Eye Res* 20:705–732.
- Kaplan HJ, Leibole MA, Tezel T, and Ferguson TA (1999) Fas ligand (CD95 ligand) controls angiogenesis beneath the retina. Nat Med 5:292– 297
- Krzystolik MG, Afshari MA, Adamis AP, et al. (2002) Prevention of experimental choroidal neovascularization with intravitreal anti-vascular endothelial growth factor antibody fragment. Arch Ophthalmol 120:338– 346.
- Okamoto N, Tobe T, Hackett SF, Ozaki H, Vinores MA, LaRochelle W, Zack DJ, and Campochiaro PA (1997) Transgenic mice with increased expression of vascular endothelial growth factor in the retina: a new model of intraretinal and subretinal neovascularization. *Am J Pathol* 151:281–291.
- Friedman DS, O'Colmain BJ, Munoz B, Tomany SC, McCarty C, de Jong PT, Nemesure B, Mitchell P, and Kempen J (2004) Prevalence of age-related macular degeneration in the United States. *Arch Ophthalmol* 122:564–572.
- Haddad S, Chen CA, Santangelo SL, and Seddon JM (2006) The genetics of age-related macular degeneration: a review of progress to date. Surv Ophthalmol 51:316–363.
- Rattner A and Nathans J (2006) Macular degeneration: recent advances and therapeutic opportunities. Nat Rev Neurosci 7:860–872.
- Bressler NM (2002) Early detection and treatment of neovascular agerelated macular degeneration. J Am Board Fam Pract 15:142–152.
- Dithmar S, Sharara NA, Curcio CA, Le NA, Zhang Y, Brown S, and Grossniklaus HE (2001) Murine high-fat diet and laser photochemical model of basal deposits in Bruch's membrane. *Arch Ophthalmol* 119:1643–1649.
- Green WR and Key SN 3rd (1977) Senile macular degeneration: a histopathologic study. Trans Am Ophthalmol Soc 75:180–254.
- Hageman GS and Mullins RF (1999) Molecular composition of drusen as related to substructural phenotype. Mol Vis 5:28.
- Kvanta A, Algvere PV, Berglin L, and Seregard S (1996) Subfoveal fibrovascular membranes in age-related macular degeneration express vascular endothelial growth factor. *Invest Ophthalmol Vis Sci* 37:1929–1934.
- Hageman GS, Anderson DH, Johnson LV, et al. (2005) A common haplotype in the complement regulatory gene factor H (HF1/CFH) predisposes individuals to age-related macular degeneration. *Proc Natl Acad Sci USA* 102:7227–7232.

- Mullins RF, Russell SR, Anderson DH, and Hageman GS (2000) Drusen associated with aging and age-related macular degeneration contain proteins common to extracellular deposits associated with atherosclerosis, elastosis, amyloidosis, and dense deposit disease. FASEB J 14:835– 846.
- Mullins RF, Aptsiauri N, and Hageman GS (2001) Structure and composition of drusen associated with glomerulonephritis: implications for the role of complement activation in drusen biogenesis. Eye 15:390–395.
- Anderson RE, Maude MB, McClellan M, Matthes MT, Yasumura D, and LaVail MM (2002) Low docosahexaenoic acid levels in rod outer segments of rats with P23H and S334ter rhodopsin mutations. *Mol Vis* 8:351–358.
- Johnson LV, Leitner WP, Staples MK, and Anderson DH (2001)
 Complement activation and inflammatory processes in Drusen formation and age related macular degeneration. Exp Eye Res 73:887–896.
- Crabb JW, Miyagi M, Gu X, et al. (2002) Drusen proteome analysis: an approach to the etiology of age-related macular degeneration. *Proc Natl Acad Sci USA* 99:14682–14687.
- Yang Z, Camp NJ, Sun H, et al. (2006) A variant of the HTRA1 gene increases susceptibility to age-related macular degeneration. Science 314:992–993
- Gehrs KM, Anderson DH, Johnson LV, and Hageman GS (2006) Agerelated macular degeneration—emerging pathogenetic and therapeutic concepts. Ann Med 38:450–471.
- Smith W, Assink J, Klein R, et al. (2001) Risk factors for age-related macular degeneration: pooled findings from three continents.
 Ophthalmology 108:697–704.
- Dhubhghaill SS, Cahill MT, Campbell M, Cassidy L, Humphries MM, and Humphries P (2010) The pathophysiology of cigarette smoking and agerelated macular degeneration. Adv Exp Med Biol 664:437–446.
- Cano M, Thimmalappula R, Fujihara M, Nagai N, Sporn M, Wang AL, Neufeld AH, Biswal S, and Handa JT (2010) Cigarette smoking, oxidative stress, the anti-oxidant response through Nrf2 signaling, and age-related macular degeneration. Vision Res 50:652–664.
- Swaroop A, Chew EY, Rickman CB, and Abecasis GR (2009) Unraveling a multifactorial late-onset disease: from genetic susceptibility to disease mechanisms for age-related macular degeneration. *Annu Rev Genomics Hum Genet* 10:19–43.
- Hirvela H, Luukinen H, Laara E, Sc L, and Laatikainen L (1996) Risk factors of age-related maculopathy in a population 70 years of age or older. *Ophthalmology* 103:871–877.
- Klein R, Klein BE, and Moss SE (1998) Relation of smoking to the incidence of age-related maculopathy. The Beaver Dam eye study. Am J Epidemiol 147:103–110.
- Seddon JM, Rosner B, Sperduto RD, Yannuzzi L, Haller JA, Blair NP, and Willett W (2001) Dietary fat and risk for advanced age-related macular degeneration. Arch Ophthalmol 119:1191–1199.
- 31. Montezuma SR, Sobrin L, and Seddon JM (2007) Review of genetics in age related macular degeneration. Semin Ophthalmol 22:229–240.
- Klein R, Klein BE, Knudtson MD, Wong TY, Cotch MF, Liu K, Burke G, Saad MF, and Jacobs DR Jr (2006) Prevalence of age-related macular degeneration in 4 racial/ethnic groups in the multi-ethnic study of atherosclerosis. Ophthalmology 113:373–380.
- Bressler SB, Munoz B, Solomon SD, and West SK (2008) Racial differences in the prevalence of age-related macular degeneration: the Salisbury Eye Evaluation (SEE) Project. Arch Ophthalmol 126:241

 –245.
- De Jong PT, Klaver CC, Wolfs RC, Assink JJ, and Hofman A (1997) Familial aggregation of age-related maculopathy. Am J Ophthalmol 124:862–863.
- Heiba IM, Elston RC, Klein BE, and Klein R (1994) Sibling correlations and segregation analysis of age-related maculopathy: the Beaver Dam eye study. Genet Epidemiol 11:51–67.
- Klaver CC, Wolfs RC, Assink JJ, van Duijn CM, Hofman A, and de Jong PT (1998) Genetic risk of age-related maculopathy. Population-based familial aggregation study. Arch Ophthalmol 116:1646–1651.

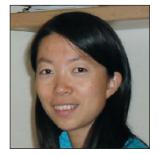
- Seddon JM, Ajani UA, and Mitchell BD (1997) Familial aggregation of age-related maculopathy. Am J Ophthalmol 123:199–206.
- Hammond CJ, Webster AR, Snieder H, Bird AC, Gilbert CE, and Spector TD (2002) Genetic influence on early age-related maculopathy: a twin study. Ophthalmology 109:730–736.
- Klein ML, Mauldin WM, and Stoumbos VD (1994) Heredity and agerelated macular degeneration. Observations in monozygotic twins. Arch Ophthalmol 112:932–937.
- Meyers SM (1994) A twin study on age-related macular degeneration. Trans Am Ophthalmol Soc 92:775–843.
- 41. Grizzard SW, Arnett D, and Haag SL (2003) Twin study of age-related macular degeneration. *Ophthalmic Epidemiol* **10**:315–322.
- Dosso AA and Bovet J (1992) Monozygotic twin brothers with age-related macular degeneration. Ophthalmologica 205:24–28.
- Klein BE, Klein R, Lee KE, Moore EL, and Danforth L (2001) Risk of incident age-related eye diseases in people with an affected sibling: The Beaver Dam Eye Study. Am J Epidemiol 154:207–211.
- Smith W and Mitchell P (1998) Family history and age-related maculopathy: the Blue Mountains Eye Study. Aust N Z J Ophthalmol 26:203–206.
- 45. Hyman LG, Lilienfeld AM, Ferris FL 3rd, and Fine SL (1983) Senile macular degeneration: a case-control study. *Am J Epidemiol* **118**:213–227.
- Seddon JM, Ajani UA, and Mitchell BD (1997) Familial aggregation of age-related maculopathy. Am J Ophthalmol 123:199–206.
- Baird PN, Guida E, Chu DT, Vu HT, and Guymer RH (2004) The epsilon2 and epsilon4 alleles of the apolipoprotein gene are associated with agerelated macular degeneration. *Invest Ophthalmol Vis Sci* 45:1311–1315.
- Klaver CC, Kliffen M, van Duijn CM, Hofman A, Cruts M, Grobbee DE, van Broeckhoven C, and de Jong PT (1998) Genetic association of apolipoprotein E with age-related macular degeneration. *Am J Hum Genet* 63:200–206.
- Schmidt S, Klaver C, Saunders A, et al. (2002) A pooled case-control study of the apolipoprotein E (APOE) gene in age-related maculopathy. Ophthalmic Genet 23:209–223.
- Souied EH, Benlian P, Amouyel P, Feingold J, Lagarde JP, Munnich A, Kaplan J, Coscas G, and Soubrane G (1998) The epsilon4 allele of the apolipoprotein E gene as a potential protective factor for exudative agerelated macular degeneration. Am J Ophthalmol 125:353–359.
- Zareparsi S, Reddick AC, Branham KE, Moore KB, Jessup L, Thoms S, Smith-Wheelock M, Yashar BM, and Swaroop A (2004) Association of apolipoprotein E alleles with susceptibility to age-related macular degeneration in a large cohort from a single center. *Invest Ophthalmol Vis Sci* 45:1306–1310.
- Stone EM, Braun TA, Russell SR, Kuehn MH, Lotery AJ, Moore PA, Eastman CG, Casavant TL, and Sheffield VC (2004) Missense variations in the fibulin 5 gene and age-related macular degeneration. N Engl J Med 351:346–353.
- Zareparsi S, Buraczynska M, Branham KE, et al. (2005) Toll-like receptor 4 variant D299G is associated with susceptibility to age-related macular degeneration. *Hum Mol Genet* 14:1449–1455.
- Yang Z, Stratton C, Francis PJ, et al. (2008) Toll-like receptor 3 and geographic atrophy in age-related macular degeneration. N Engl J Med 359:1456–1463.
- Neale BM, Fagerness J, Reynolds R, et al. (2010) Genome-wide association study of advanced age-related macular degeneration identifies a role of the hepatic lipase gene (LIPC). Proc Natl Acad Sci USA 107:7395–7400.
- Chen W, Stambolian D, Edwards AO, et al. (2010) Genetic variants near TIMP3 and high-density lipoprotein-associated loci influence susceptibility to age-related macular degeneration. *Proc Natl Acad Sci USA* 107:7401–7406
- Edwards AO, Ritter R 3rd, Abel KJ, Manning A, Panhuysen C, and Farrer LA (2005) Complement factor H polymorphism and age-related macular degeneration. Science 308:421–424.

- Zareparsi S, Branham KE, Li M, Shah S, Klein RJ, Ott J, Hoh J, Abecasis GR, and Swaroop A (2005) Strong association of the Y402H variant in complement factor H at 1q32 with susceptibility to age-related macular degeneration. Am J Hum Genet 77:149–153.
- Gold B, Merriam JE, Zernant J, et al. (2006) Variation in factor B (BF) and complement component 2 (C2) genes is associated with age-related macular degeneration. *Nat Genet* 38:458–462.
- Li M, Atmaca-Sonmez P, Othman M, et al. (2006) CFH haplotypes without the Y402H coding variant show strong association with susceptibility to age-related macular degeneration. Nat Genet 38:1049–1054.
- Maller J, George S, Purcell S, Fagerness J, Altshuler D, Daly MJ, and Seddon JM (2006) Common variation in three genes, including a noncoding variant in CFH, strongly influences risk of age-related macular degeneration. *Nat Genet* 38:1055–1059.
- Fisher SA, Abecasis GR, Yashar BM, et al. (2005) Meta-analysis of genome scans of age-related macular degeneration. *Hum Mol Genet* 14:2257–2264
- Jakobsdottir J, Conley YP, Weeks DE, Mah TS, Ferrell RE, and Gorin MB (2005) Susceptibility genes for age-related maculopathy on chromosome 10q26. Am J Hum Genet 77:389–407.
- Rivera A, Fisher SA, Fritsche LG, Keilhauer CN, Lichtner P, Meitinger T, and Weber BH (2005) Hypothetical LOC387715 is a second major susceptibility gene for age-related macular degeneration, contributing independently of complement factor H to disease risk. *Hum Mol Genet* 14:3227–3236.
- Schmidt S, Hauser MA, Scott WK, et al. (2006) Cigarette smoking strongly modifies the association of LOC387715 and age-related macular degeneration. Am J Hum Genet 78:852–864.
- Dewan A, Liu M, Hartman S, et al. (2006) HTRA1 promoter polymorphism in wet age-related macular degeneration. Science 314:989–992.
- Pangburn MK, Pangburn KL, Koistinen V, Meri S, and Sharma AK (2000) Molecular mechanisms of target recognition in an innate immune system: interactions among factor H, C3b, and target in the alternative pathway of human complement. *J Immunol* 164:4742–4751.
- Ranganathan S, Male DA, Ormsby RJ, Giannakis E, and Gordon DL (2000) Pinpointing the putative heparin/sialic acid-binding residues in the 'sushi' domain 7 of factor H: a molecular modeling study. Pac Symp Biocomput 2000:155–167.
- Tocharus J, Tsuchiya A, Kajikawa M, Ueta Y, Oka C, and Kawaichi M (2004) Developmentally regulated expression of mouse HtrA3 and its role as an inhibitor of TGF-beta signaling. Dev Growth Differ 46:257–274.
- Grau S, Richards PJ, Kerr B, et al. (2006) The role of human HtrA1 in arthritic disease. J Biol Chem 281:6124-6129.
- Oka C, Tsujimoto R, Kajikawa M, et al. (2004) HtrA1 serine protease inhibits signaling mediated by TGFβ family proteins. *Development* 131:1041–1053
- Haines JL, Hauser MA, Schmidt S, et al. (2005) Complement factor H variant increases the risk of age-related macular degeneration. *Science* 308:419–421.
- Klein RJ, Zeiss C, Chew EY, et al. (2005) Complement factor H polymorphism in age-related macular degeneration. Science 308:385–389.
- Maller JB, Fagerness JA, Reynolds RC, Neale BM, Daly MJ, and Seddon JM (2007) Variation in complement factor 3 is associated with risk of age-related macular degeneration. *Nat Genet* 39:1200–1201.
- Yates JR, Sepp T, Matharu BK, et al. (2007) Complement C3 variant and the risk of age-related macular degeneration. N Engl J Med 357:553– 561
- Seddon JM, Reynolds R, Maller J, Fagerness JA, Daly MJ, and Rosner B (2009) Prediction model for prevalence and incidence of advanced age-related macular degeneration based on genetic, demographic, and environmental variables. *Invest Ophthalmol Vis Sci* 50:2044–2053.
- Seitsonen SP, Onkamo P, Peng G, et al. (2008) Multifactor effects and evidence of potential interaction between complement factor H Y402H and LOC387715 A69S in age-related macular degeneration. *PLoS One* 3:e3833.

- Schaumberg DA, Hankinson SE, Guo Q, Rimm E, and Hunter DJ (2007) A prospective study of 2 major age-related macular degeneration susceptibility alleles and interactions with modifiable risk factors. *Arch Ophthalmol* 125:55–62.
- Swaroop A, Branham KE, Chen W, and Abecasis G (2007) Genetic susceptibility to age-related macular degeneration: a paradigm for dissecting complex disease traits. *Hum Mol Genet* 16:R174–182.
- Reynolds R, Hartnett ME, Atkinson JP, Giclas PC, Rosner B, and Seddon JM (2009) Plasma complement components and activation fragments: associations with age-related macular degeneration genotypes and phenotypes. *Invest Ophthalmol Vis Sci* 50:5818–5827.
- Scholl HP, Charbel Issa P, Walier M, et al. (2008) Systemic complement activation in age-related macular degeneration. PLoS One 3:e2593.
- Brantley MA Jr, Edelstein SL, King JM, Plotzke MR, Apte RS, Kymes SM, and Shiels A (2009) Association of complement factor H and LOC387715 genotypes with response of exudative age-related macular degeneration to photodynamic therapy. Eye (Lond) 23:626–631.
- 83. Goverdhan SV, Hannan S, Newsom RB, Luff AJ, Griffiths H, and Lotery AJ (2008) An analysis of the CFH Y402H genotype in AMD patients and controls from the UK, and response to PDT treatment. Eye (Lond) 22:849–854.
- Seitsonen SP, Jarvela IE, Meri S, Tommila PV, Ranta PH, and Immonen IJ (2007) The effect of complement factor H Y402H polymorphism on the outcome of photodynamic therapy in age-related macular degeneration. *Eur J Ophthalmol* 17:943–949.
- Feng X, Xiao J, Longville B, et al. (2009) Complement factor H Y402H and C-reactive protein polymorphism and photodynamic therapy response in age-related macular degeneration. *Ophthalmology* 116:1908–1912 e1901.
- Chowers I, Meir T, Lederman M, et al. (2008) Sequence variants in HTRA1 and LOC387715/ARMS2 and phenotype and response to photodynamic therapy in neovascular age-related macular degeneration in populations from Israel. Mol Vis 14:2263–2271.
- Immonen I, Seitsonen S, Tommila P, Kangas-Kontio T, Kakko S, Savolainen ER, Savolainen MJ, and Liinamaa MJ (2010) Vascular endothelial growth factor gene variation and the response to photodynamic therapy in age-related macular degeneration. *Ophthalmology* 117:103–108.
- Parmeggiani F, Costagliola C, Gemmati D, et al. (2007) Predictive role of coagulation-balance gene polymorphisms in the efficacy of photodynamic therapy with verteporfin for classic choroidal neovascularization secondary to age-related macular degeneration. *Pharmacogenet Genomics* 17:1039–1046.
- Parmeggiani F, Costagliola C, Gemmati D, et al. (2008) Coagulation gene predictors of photodynamic therapy for occult choroidal neovascularization in age-related macular degeneration. *Invest Ophthalmol Vis Sci* 49:3100–3106.
- Parmeggiani F, Gemmati D, Costagliola C, Sebastiani A, and Incorvaia C (2009) Predictive role of C677T MTHFR polymorphism in variable efficacy of photodynamic therapy for neovascular age-related macular degeneration. *Pharmacogenomics* 10:81–95.
- Brown DM, Kaiser PK, Michels M, Soubrane G, Heier JS, Kim RY, Sy JP, and Schneider S (2006) Ranibizumab versus verteporfin for neovascular age-related macular degeneration. N Engl J Med 355:1432-1444.
- Rosenfeld PJ, Brown DM, Heier JS, Boyer DS, Kaiser PK, Chung CY, and Kim RY (2006) Ranibizumab for neovascular age-related macular degeneration. N Engl J Med 355:1419–1431.
- Avery RL, Pieramici DJ, Rabena MD, Castellarin AA, Nasir MA, and Giust MJ (2006) Intravitreal bevacizumab (Avastin) for neovascular agerelated macular degeneration. Ophthalmology 113:363–372.e5.
- Brantley MA Jr, Fang AM, King JM, Tewari A, Kymes SM, and Shiels A (2007) Association of complement factor H and LOC387715 genotypes with response of exudative age-related macular degeneration to intravitreal bevacizumab. *Ophthalmology* 114:2168–2173.

- Lee AY, Raya AK, Kymes SM, Shiels A, and Brantley MA Jr (2009)
 Pharmacogenetics of complement factor H (Y402H) and treatment of exudative age-related macular degeneration with ranibizumab. Br J Ophthalmol 93:610–613.
- Clemons TE, Milton RC, Klein R, Seddon JM, and Ferris FL 3rd (2005)
 Risk factors for the incidence of advanced age-related macular degeneration in the age-related eye disease study (AREDS): AREDS report no.
 19. Ophthalmology 112:533–539.
- Anonymous (2001) A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss: AREDS report no 8. Arch *Ophthalmol* 119:1417–1436.
- Klein ML, Francis PJ, Rosner B, Reynolds R, Hamon SC, Schultz DW, Ott J, and Seddon JM (2008) CFH and LOC387715/ARMS2 genotypes and treatment with antioxidants and zinc for age-related macular degeneration. *Ophthalmology* 115:1019–1025.
- Link E, Parish S, Armitage J, Bowman L, Heath S, Matsuda F, Gut I, Lathrop M, and Collins R (2008) SLCO1B1 variants and statin-induced myopathy--a genomewide study. N Engl J Med 359:789–799.
- Druker BJ, Guilhot F, O'Brien SG, et al. (2006) Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. N Engl J Med 355:2408–2417.
- 101. Aithal GP, Day CP, Kesteven PJ, and Daly AK (1999) Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 353:717–719.
- 102. Allikmets R (2000) Further evidence for an association of ABCR alleles with age-related macular degeneration. The International ABCR Screening Consortium. Am J Hum Genet 67:487–491.
- 103. Katta S, Kaur I, and Chakrabarti S (2009) The molecular genetic basis of age-related macular degeneration: an overview. J Genet 88:425–449.
- 104. Thakkinstian A, Bowe S, McEvoy M, Smith W, and Attia J (2006) Association between apolipoprotein E polymorphisms and age-related macular degeneration: A HuGE review and meta-analysis. Am J Epidemiol 164:813–822.
- 105. Francis PJ, Hamon SC, Ott J, Weleber RG, and Klein ML (2009) Polymorphisms in C2, CFB and C3 are associated with progression to advanced age related macular degeneration associated with visual loss. J Med Genet 46:300–307.
- 106. Ross RJ, Bojanowski CM, Wang JJ, Chew EY, Rochtchina E, Ferris FL 3rd, Mitchell P, Chan CC, and Tuo J (2007) The LOC387715 polymorphism and age-related macular degeneration: replication in three case-control samples. *Invest Ophthalmol Vis Sci* 48:1128–1132.
- 107. Chen W, Xu W, Tao Q, Liu J, Li X, Gan X, Hu H, and Lu Y (2009) Metaanalysis of the association of the HTRA1 polymorphisms with the risk of age-related macular degeneration. Exp Eye Res 89:292–300.
- 108. Tang NP, Zhou B, Wang B, and Yu RB (2009) HTRA1 promoter polymorphism and risk of age-related macular degeneration: a meta-analysis. Ann Epidemiol 19:740–745.
- 109. Thakkinstian A, Han P, McEvoy M, Smith W, Hoh J, Magnusson K, Zhang K, and Attia J (2006) Systematic review and meta-analysis of the association between complement factor H Y402H polymorphisms and age-related macular degeneration. *Hum Mol Genet* 15:2784–2790.
- 110. Spencer KL, Hauser MA, Olson LM, Schmidt S, Scott WK, Gallins P, Agarwal A, Postel EA, Pericak-Vance MA, and Haines JL (2008) Deletion of CFHR3 and CFHR1 genes in age-related macular degeneration. *Hum Mol Genet* 17:971–977.
- 111. Combadiere C, Feumi C, Raoul W, et al. (2007) CX3CR1-dependent subretinal microglia cell accumulation is associated with cardinal features of age-related macular degeneration. J Clin Invest 117:2920–2928.
- 112. Ding X, Patel M, and Chan CC (2009) Molecular pathology of age-related macular degeneration. *Prog Retin Eye Res* 28:1–18.
- 113. Tuo J, Ning B, Bojanowski CM, et al. (2006) Synergic effect of polymorphisms in ERCC6 5' flanking region and complement factor H on agerelated macular degeneration predisposition. *Proc Natl Acad Sci USA* 103:9256–9261.

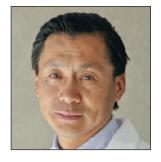
- 114. Edwards AO, Chen D, Fridley BL, et al. (2008) Toll-like receptor polymorphisms and age-related macular degeneration. *Invest Ophthalmol Vis Sci* 49:1652–1659.
- 115. Cho Y, Wang JJ, Chew EY, Ferris FL 3rd, Mitchell P, Chan CC, and Tuo J (2009) Toll-like receptor polymorphisms and age-related macular degeneration: replication in three case-control samples. *Invest Ophthalmol Vis Sci* 50:5614–5618
- 116. Galan A, Ferlin A, Caretti L, Buson G, Sato G, Frigo AC, and Foresta C (2010) Association of age-related macular degeneration with polymorphisms in vascular endothelial growth factor and its receptor. Ophthalmology 117:1769–1774.
- 117. Churchill AJ, Carter JG, Lovell HC, Ramsden C, Turner SJ, Yeung A, Escardo J, and Atan D (2006) VEGF polymorphisms are associated with neovascular age-related macular degeneration. *Hum Mol Genet* 15:2055–2061
- 118. Fang AM, Lee AY, Kulkarni M, Osborn MP, and Brantley MA Jr (2009) Polymorphisms in the VEGFA and VEGFR-2 genes and neovascular age-related macular degeneration. Mol Vis 15:2710–2719.
- 119. Boekhoorn SS, Isaacs A, Uitterlinden AG, van Duijn CM, Hofman A, de Jong PT, and Vingerling JR (2008) Polymorphisms in the vascular endothelial growth factor gene and risk of age-related macular degeneration: the Rotterdam Study. *Ophthalmology* 115:1899–1903.



Yuhong Chen, MD, PhD, is an ophthalmologist at the Eye and ENT Hospital of Fudan University in China. She has been working as a postdoctoral fellow in ocular genetics at UC San Diego since 2008.



Matthew Bedell, MD, graduated this year from the UCSD School of Medicine. He is working as a post-doctoral scholar in ocular genetics and planning on a career in clinical and research ophthalmology.



Kang Zhang, MD, PhD, is a faculty member at Shiley Eye Center and director of the Institute for Genomic Medicine at UCSD. His research mainly focuses on novel disease gene targets and therapies for macular degeneration, glaucoma, diabetic retinopathy, and inherited retinal degenerations. Address correspondence to KZ.

E-mail kang.zhang@gmail.com; fax 858-246-0873.